Use of Simulation to Promote Best Practice for Obtaining Blood Cultures

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Kentucky Infection Prevention Training Center

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Project Firstline is a national collaborative led by the U.S. Centers for Disease Control and Prevention (CDC) to provide infection control training and education to frontline health care workers and public health personnel. Kentucky Infection Prevention (KyIP) Training Center is proud to partner with Project Firstline to deliver the most up-to-date and best quality infection prevention and control training and information.



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Project Firstline



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Objectives

- Define what a blood culture is and how it is used for diagnostic and treatment planning.
- Recognize the impact of blood culture contamination on patients and the healthcare system as a whole.
- Analyze the relationship between blood culture contamination and antibiotic stewardship.
- Apply actions that will prevent blood culture contamination.
- Demonstrate proper method for blood culture collection in regards to technique, volume, and timing.



Risk Recognition





Kentucky Infection Prevention Training Center

Pre-Brief (Background)

- Episode 21: Do We Really Have to Talk About Hand Hygiene? Again? Yes!
- Project Firstline, CDC
- Dr. Abby Carlson, MD





Reservoirs

- Where germs can commonly live in and on the body
 - Skin
 - Gut
 - Respiratory
 - Blood





What is a blood culture?

- A sample of blood drawn from the patient that is placed in media and monitored for growth.
- Growth of bacteria could indicate the patient has bacteria in their bloodstream.
 - Contamination of a blood culture could falsely indicate a bloodstream infection
- Blood cultures are important diagnostic tools for patients presenting with signs of bacteremia
- Determining organism type via culture guides treatment and management





How do blood cultures guide patient management?

- If an organism is detected, the most appropriate antibiotic is selected to treat that organism.
- Some organisms are always clinically significant, but others can likely be a blood culture contamination.
- The provider will compare the patient's clinical presentation if a contamination is suspected.

Always clinically significant	Typically contaminant, but should check for
	clinical correlation
S. aureus	Coagulase-negative staphylococci (except
	Staphylococcus lugdunensis, this is typically
	clinically significant)
Streptococcus pneumoniae	Corynebacterium species
Group A Streptococcus	Cutibacterium acnes
Enterobacteriaceae	Bacillus species (except B. anthracis, which is
	a clinically significant pathogen)
Haemophilus influenzae	Micrococcus species
Pseudomonas aeruginosa	
Bacteroidaceae	
Candida species	



(Doern, 2021)

Impact of Blood Culture Contamination

- Longer hospital stays
- Unnecessary antibiotic use
- Unnecessary antibiotic use can lead to antibiotic resistance



How do blood cultures get contaminated?

- At the time of the venipuncture
 - Inadequate skin antisepsis
- During transfer to the blood culture bottle
 - Inadequate disinfection of rubber diaphragm of blood culture bottle
- Indwelling lines





Germs Live on Skin









Germs Live on Dry Surfaces









Prevention of Contamination

- Careful ordering of blood cultures based on appropriate patient selection
- Skin antisepsis with 2% alcoholic chlorohexidine or 70% isopropyl alcohol, followed by 2% chlorhexidine. Allow at least 30 seconds for dry time.
- Disinfect the rubber diaphragm of the blood culture vials with at least 70% isopropyl alcohol.
- Blood cultures should routinely be collected from peripheral sites, not central lines or other intravascular access sites which increase the chance of contamination.
- Generally hand hygiene with clean nonsterile gloves are adequate and recommended. Sterile gloves should be used if re-palpation of the disinfected skin site in necessary.
- Blood culture kits and standard procedures for obtaining cultures have shown in studies to reduce contamination rates.



Prevention of Contamination

- When obtaining multiple labs during one venipuncture, blood cultures should be drawn first to prevent contamination.
- Blood cultures should be drawn directly into blood culture bottles using a butterfly and an adapter whenever possible. This process decreases the risk of blood culture contamination through the transfer process.



Blood Culture Contamination Rates

- American Society for Microbiology (ASM) and Clinical Laboratory Standards Institute (CLSI), blood culture contamination rates should not exceed 3%
- However, per the CDC, a contamination rate of less than 1% is achievable when best practices are in place. The CDC emphasizes striving for <1%.
 - Proper hand hygiene
 - Proper skin antisepsis
 - Scrubbing vial diaphragms with alcohol adequately
- Do you know your facility's current contamination rate?



Antibiotic Resistance and Stewardship

- "Clinicians should strive to obtain blood cultures for the right patients, in the right settings, and at the right time," (CDC, n.d. pg. 2).
- Starting antibiotics without obtaining blood cultures first makes narrowing down antibiotics much harder
- Ordering blood cultures on a patient who has a low likelihood of bacteremia could end up with a false positive result
- Goal of antibiotic stewardship is to reduce the use of antibiotics, therefore reducing resistance





Blood Volume

• In adults obtain 20mls (10ml for aerobic and 10ml for anaerobic) for the most accurate pathogen yield.





Dos	Don'ts



Dos	Don'ts
DO order to determine etiology of infection	DON'T order blood cultures routinely on every patient



Dos	Don'ts
DO order to determine etiology of infection	DON'T order blood cultures routinely on every
DO obtain cultures before starting antibiotics	patient
	DON'T overfill or under fill blood culture bottles



Dos	Don'ts
DO order to determine etiology of infection	DON'T order blood cultures routinely on every
DO obtain cultures before starting antibiotics	patient
DO use proper skin antisepsis technique	DON'T overfill or under fill blood culture bottles
	DON'T use alcohol only, scrubbing back and forth



Dos	Don'ts
DO order to determine etiology of infection	DON'T order blood cultures routinely on every
DO obtain cultures before starting antibiotics	patient
DO use proper skin antisepsis technique	DON'T overfill or under fill blood culture bottles
DO choose peripheral sites over indwelling lines	DON'T use alcohol only, scrubbing back and forth
Do choose peripheral sites over indivening intes	DON'T draw from indwelling lines without clinician order



Dos	Don'ts
DO order to determine etiology of infection	DON'T order blood cultures routinely on every
DO obtain cultures before starting antibiotics	patient
DO use proper skin antisepsis technique	DON'T overfill or under fill blood culture bottles
	DON'T use alcohol only, scrubbing back and forth
DO choose peripheral sites over indweiling lines	DON'T draw from indwelling lines with clinician order
DO disinfect the rubber diaphragm of the vial	
	DON I retrigerate or freeze blood cultures



Dos	Don'ts
DO order to determine etiology of infection	DON'T order blood cultures routinely on every
DO obtain cultures before starting antibiotics	patient
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	DON'T use alcohol only, scrubbing back and forth
DO choose peripheral sites over indwelling lines	DON'T draw from indwolling lines with clinician order
DO disinfect the rubber diaphragm of the vial	DON T draw nom indweining imes with clinician order
DO develop a team for blood culture collection	DON'T refrigerate or freeze blood cultures
	DON'T allow your facility contamination rate >1%



Dos	Don'ts
DO order to determine etiology of infection	DON'T order blood cultures routinely on every
DO obtain cultures before starting antibiotics	patient
DO use proper skin antisensis technique	DON'T overfill or under fill blood culture bottles
	DON'T use alcohol only, scrubbing back and forth
DO choose peripheral sites over indwelling lines	DON'T draw from indwelling lines with clinician order
DO disinfect the rubber diaphragm of the vial	
DO develop a team for blood culture collection	DON'I refrigerate or freeze blood cultures
DO graata a standardized process	DON'T allow your facility contamination rate >1%
DO create a stanuaruizeu process	DON'T draw multiple blood culture vials (beyond one anaerobic and one aerobic) from the same site



Dos	Don'ts
DO order to determine etiology of infection	DON'T order blood cultures routinely on every
DO obtain cultures before starting antibiotics	patient
DO use proper skin antisepsis technique	DON'T overfill or under fill blood culture bottles
DO choose peripheral sites over indwelling lines	DON'T use alcohol only, scrubbing back and forth
DO choose periprieral sites over indivening intes	DON'T draw from indwelling lines with clinician order
DO disinfect the rubber diaphragm of the viai	DON'T refrigerate or freeze blood cultures
DO develop a team for blood culture collection	DON'T allow your facility contamination rate >1%
DO create a standardized process	
DO provide surveillance and feedback on	DON'T draw multiple blood culture vials (beyond one anaerobic and one aerobic) from the same site
contamination rates	DON'T forget to compare potential contaminated organisms to clinical presentation



Dos	Don'ts
DO order to determine etiology of infection	DON'T order blood cultures routinely on every
DO obtain cultures before starting antibiotics	patient
DO use proper skin antisepsis technique	DON'T overfill or under fill blood culture bottles
DO choose peripheral sites over indwelling lines	DON'T use alcohol only, scrubbing back and forth
DO disinfect the rubber diaphragm of the vial	DON'T draw from indwelling lines with clinician order
DO develop a team for blood culture collection	DON'T refrigerate or freeze blood cultures
DO create a standardized process	DON'T allow your facility contamination rate >1%
DO provide surveillance and feedback on	DON'T draw multiple blood culture vials (beyond
contamination rates	one anaerobic and one aerobic) from the same site
DO draw blood directly into the blood culture bottle using an initial diversion device if available	DON'T forget to compare potential contaminated organisms to clinical presentation
bottle doing an initial diversion device il available	



Dos	Don'ts
DO order to determine etiology of infection	DON'T order blood cultures routinely on every
DO obtain cultures before starting antibiotics	patient
DO use proper skin antisepsis technique	DON'T overfull or under full blood culture bottles
DO choose peripheral sites over indwelling lines	DON'T use alcohol only, scrubbing back and forth
DO disinfect the rubber diaphragm of the vial	DON'T draw from indwelling lines with clinician order
DO develop a team for blood culture collection	DON'T refrigerate or freeze blood cultures
DO create a standardized process	DON'T allow your facility contamination rate >1%
DO provide surveillance and feedback on	DON'T draw multiple blood culture vials (beyond one anaerobic and one aerobic) from the same site
DO draw blood directly into the blood culture	DON'T forget to compare potential contaminated organisms to clinical presentation
DO obtain adequate blood volume	



Indwelling verse peripheral



- Blood cultures should always be drawn peripherally from a fresh venipuncture. This is best practice.
- Drawing blood cultures off of existing central and peripheral lines can increase the risk of blood culture contamination.



• It is best practice to only draw peripheral cultures unless ordered otherwise by a provider. <u>Sometimes a provider will request cultures be drawn off of a line if a different type of infection is suspected</u>.



Simulation #1: Skin Antisepsis

Click here to

watch

- Demonstrate obtaining blood cultures from three different sites with proper and improper skin antisepsis to show contamination risks.
- Proper skin antisepsis should consist of a 2% CHG/70% alcohol prep sponge scrub for 30 seconds of back and forth vigorous scrubbing





Simulation #2: Improper Scrubbing Technique

- Demonstrate effects of proper vs improper scrubbing of rubber diaphragm of blood culture bottle
- Best practice is to disinfect the rubber diaphragm of the blood culture vials with 70% alcohol sterile pad. Allow diaphragm to dry before transfer.







Simulation #2 Activity

1.	Blood cultures routinely drawn from peripheral sites.				
2.	Proper Technique Improper Technique Using CHG/alcohol sponge, scrub the venipuncture site using back and forth motions for 30 seconds. Allow site to dry for a minimum of 30 seconds.		5.	Using 70% isopropyl alcohol pad to disinfect the rubber diaphragm of the blood culture vial and allow to dry.	
				Proper Technique	Improper Technique
3.	Proper Technique	Improper Technique	6.	Holding on to the end of the syringe as you connect the transfer device to ensure a good connection.	
	Repalpating the site wearing clean non-sterile gloves after skin antisepsis is performed.			Proper Technique	Improper Technique
	Proper Technique	Improper Technique	7.	If unable to get specimen to lab immedi transport.	iately after collection, place in freezer until
4.	Obtaining at least 10ml of blood per culture bottle.			Proper Technique	Improper Technique
	Proper Technique	Improper Technique			



Simulation #3: Blood Volume

- Visualize proper and improper blood volumes in both syringes and blood culture vials
- Discuss the relationship of blood volume and pathogen growth yield.







Simulation #3: Blood Volume





Simulation #4: Contamination Rates

 Scenario: Your facility's blood culture contamination rate is now 5%, which is up from the normal 1% contamination rate the facility usually sees. This contamination rate changed in one month's time. The instructor will test the learner's knowledge on what level of contamination is acceptable.





Simulation #4 Discussion

- The CDC emphasizes with the proper techniques and protocols that a <1% contamination rate is doable.
- Blood culture contamination can result in unnecessary antibiotic use until repeat cultures can grow.
- Obtaining blood cultures before starting antibiotics is key to finding the best antibiotic to treat the pathogen. Starting broad spectrum without cultures can make it difficult to narrow and ultimately increase the risk of antibiotic resistance.
- Knowing the dos and don'ts of blood culture collection will help keep contamination rates down.
- A standardized approach, the use of kits, and a dedicated team of people that draws the blood cultures are all ways to decrease the contamination rate.



Blood Culture Collection-Proper Method

 Steps followed as provided by Elsevier







De-Brief

- What errors did you identify?
- What is the impact?
- What practices may prevent this issue?
- How far up the chain will you take this?
- How can Human Factors Engineering be applied to this issue?
- What went well?
- What would you change?



De-Brief: Key Take-Home Points

- Hand hygiene is vital to preventing blood culture contamination. Sterile gloves are required if re-palpating the site.
- Skin is a reservoir for germs. Skin antisepsis with 2% alcoholic chlorohexidine or 70% isopropyl alcohol, followed by 2% chlorhexidine can prevent blood culture contamination.
- Disinfect the rubber diaphragm of the blood culture vials with at least 70% isopropyl alcohol.
- Choose peripheral sites for collection over indwelling lines unless instructed by a provider.
- Adequate blood volume is imperative for accurate pathogen yield.
- Contamination rates below 1% can decrease the risk of antibiotic resistance due to unnecessary antibiotic use.



Kentucky Antimicrobial Stewardship Innovation Consortium (KASIC)

- "A free program to address the use of antimicrobials across the Commonwealth in order to optimize clinical outcomes while minimizing adverse consequences of antimicrobial use."
- Kentucky prescribes antibiotics at one of the highest rates in the country. KASIC wants to change this.
- KASIC offers expert resources for education and mentorship to facilities and organizations dedicated to antibiotic stewardship.





KASIC Office Hours

- Every Thursday from 12:00-1:00pm EST
- Scan the QR code to add to your calendar
- For healthcare professionals to drop in for questions, guidance, or facility concerns.





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