Using T2 Technology to Move the Needle on Sepsis

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Learning Objectives

- Describe the principle of operation of the T2 Biosystems Platform
- List the yeast and bacterial targets on the current test menu
- Describe the optimal utilization of T2 technology in support of Diagnostic Stewardship



The ERA of Rapid Diagnostics

Why Rapid Diagnostics?

- Need for accurate and early diagnosis (culture independent; direct specimen testing)
- Culture (Gold Standard): low sensitivity, increased TAT
- Initiate appropriate therapy to reduce morbidity and mortality
- Promote and Support Antimicrobial Stewardship: empiric, targeted or discontinuation of therapy
- Reduce unnecessary use of antibiotics/antifungals, poor outcomes, length of stay and/or readmission rates related to the initial hospitalization
- Support Sepsis Program



Time to Appropriate Therapy is a Key Driver of Clinical Outcomes

- Extended time from presentation to appropriate therapy remains a <u>major</u> contributor to <u>poor patient outcomes</u> and proliferation of <u>antimicrobial resistance</u>
- Current standard of care requires up to 3 days to provide species ID and susceptibility results
- For <u>every hour delay in time to appropriate therapy</u> survival decreases by 7.6% during septic shock¹
- As many as 80% of sepsis deaths could be prevented with rapid diagnosis and treatment¹
- Prolonged use of <u>broad-spectrum antimicrobials</u> is a known risk factor associated with the development and spread of <u>antimicrobial-resistant organisms</u>



Culture Independent Tests are Simple to Use

Fully-automated T2Dx Instrument

- Rapid: results in 3 to 5 hours
- Simple: no sample preparation
- Ultra-sensitive: as low as 1 CFU/mL
- T2MR[®] technology is not inhibited by prior antimicrobial administration¹



T2Candida®

Sensitivity: 91%² Specificity: 99%²

- C. albicans
- C. tropicalis
- C. parapsilosis
- C. krusei
- C. glabrata

FDA-Cleared CE marked 1-3 CFU/mL LoD

T2 Bacteria®

Sensitivity: 90%³ Specificity: 98%³

- E. faecium
- S. aureus
- K. pneumoniae
- A. baumannii
- P. aeruginosa
- E. coli

FDA-Cleared CE marked 2-11 CFU/mL LoD

T2 Resistance

FDA Breakthrough Device CEmark/RUO 2019

mecA/C

vanA/B

CTXM-14/15

KPC

OXA-48 Group

NDM, VIM, IMP

AmpC (CMY/DHA)

U.S. Clinical Trial

(patient enrollment complete)

CE marked

3-11 CFU/mL LoD

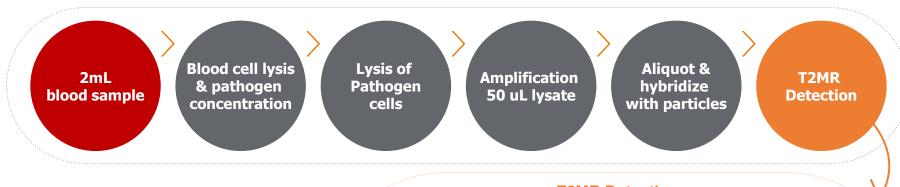


SCHOOL OF MEDICINE

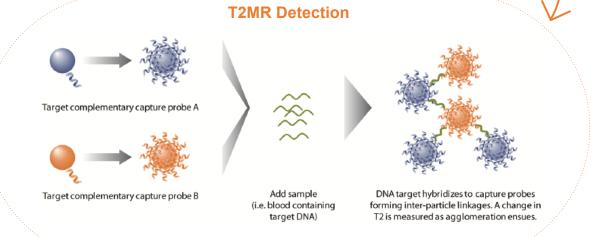
DEPARTMENT OF PATHOLOGY & LABORATORY MEDICINE

- 1. T2Candida and T2Bacteria Instructions for Use, refer to Performance Characteristics: Interfering Substances
- 2. Mylonakis, E., Clancy, C.J., Ostrosky-Zeichner, L., et al. (2015). Clinical Infectious Diseases
- 3. Nguyen H, et al. Performance of the T2Bacteria Panel for Diagnosing Bloodstream Infections: A Diagnostic Accuracy Study. Ann Intern Med. 2019.

T2MR Direct Detection for Superior Results

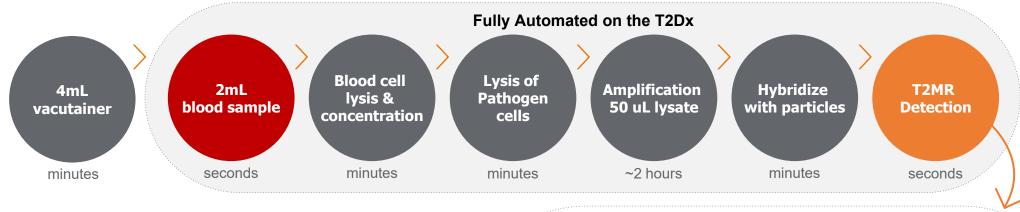


- T2 Magnetic Resonance (T2MR) enables measurement direct from patient sample enabling higher sensitivity
- Unparalleled 1 CFU/ml sensitivity in complex matrixes
- No background interference of T2MR signals eliminates sample preparation and extraction of targets

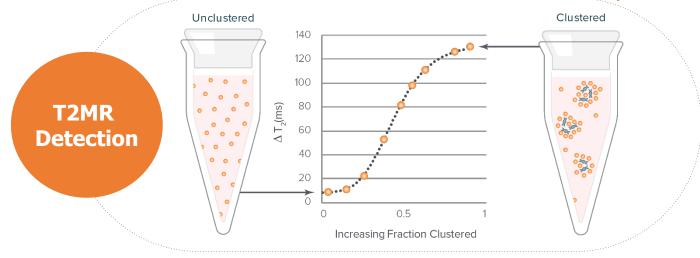




T2Dx Instrument Workflow

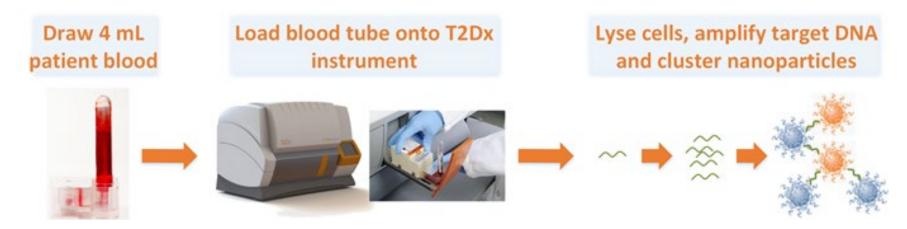


Measuring the magnetic properties of the entire water population and not just the target provides breakthrough sensitivity in complex clinical samples





Workflow



- Proprietary methodology enabling inhibition-free DNA amplification in complex clinical matrices.
 Utilizes miniaturized magnetic resonance technology which measures how water molecules react in presence of magnetic field.
- No background interference (e.g. human DNA, antibiotics, etc.) simplifies process and eliminates extraction & purification of targets
- T2 Magnetic Resonance (T2MR) enables measurement direct from patient sample enables higher sensitivity.

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T2SEPSIS SOLUTION™

SAMPLE COLLECTION PRACTICE FOR OPTIMAL PERFORMANCE

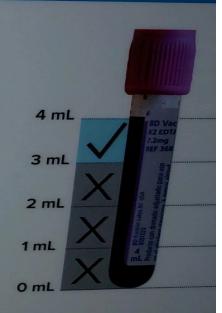
SAMPLE COLLECTION

- Decontaminate the top of a 4 mL K₂ED N purple/lavender-top Vacutainer and allow to dry.
- · Draw blood sample into the decontaminated 4 mL Vacutainer using the same aseptic technique and anatomical location as the blood culture draw.

NOTE: SAMPLE WILL BE REJECTED IF VOLUME IS NOT AT LEAST 3 ML

- Invert the filled vacutainer 8-10 times following draw to properly mix the sample and K2EDTA.
- Send the sample to the laboratory for processing as soon as possible. The sample may be stored at 15-25°C for no longer than 12 hours before processing.

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Must be accompanied by a peripheral blood culture collected within one hour



Invalid Results

Understanding the Results Report

Sample Check

- ✓ Volume of \ge 3 mL in a 4 mL K₂EDTA
- ✓ Stored properly
 - 15-25°C ≤ 12 hours
 - 2-8°C ≤ 24 hours
- Equilibrated to room temperature prior to Panel assembly
- Check for clots

Procedure Check

- Only use 4 mL K₂EDTA tubes (lavender top)
- Use aseptic draw technique for sample collection
- ✓ Invert sample tube 8-10x prior to Panel assembly
- Sample Inlet "clicks" when attached to Cartridge
- ✓ Blood fills both wells of Sample Inlet
- Reagent tray vortexed at 2560-3200
 RPM and flicked
- ✓ Temperature between 20-25°C

Interferent Check*

- ✓ Calcium Hypochlorite (20 mg/mL)
- \checkmark K₂EDTA (≥ 3.0 mg/mL)
- ✓ Feraheme (≥ 76.5 mg/mL)
- ✓ MRI Contrast Agents
 - Magnevist (≥ 1.7 mg/mL)
 - Ablavar (≥ 0.39 mg/mL)
- ✓ Intralipid (≥ 160 mg/dL)
 - Lipemia
 - TPN
 - Lipid Emulsion



^{*}The effect of interfering substances has only been evaluated for those listed in the labeling. Interference by substances other than those described in the Interference section could lead to erroneous results.

Promote Optimal Utilization (Diagnostic Stewardship)

- Testing restricted to high-risk patients:
 - Intensive Care Units
 - Transplant Patients (BMT and Solid Organ)
 - Oncology
 - Emergency Room
 - Infectious Diseases
- Training of phlebotomists, nursing, and clinicians
 - T2 sample collection (3 4ml in separate tube); peripheral only, no line draws
 - Blood culture collected simultaneously (20 ml: aerobic/anaerobic)



Ordering Criteria and Establishing Order Set

- Testing restricted to High Risk Patients:
 - Intensive Care Units: recommend T2Bacteria Panel
 - Transplant (BMT and Solid Organ): recommend T2Candida Panel
 - Oncology: recommend T2Candida Panel
 - Emergency Room: recommend T2Bacteria Panel
 - Infectious Diseases: T2Candida or T2Bacteria Panel
- Training of phlebotomists and nursing personnel: specimen collection (T2 sample and Blood Culture): Critical!



Results

	T2 Assay n=5	Control n=5	p-value
Primary outcome Time to therapy, median (days)	0.3	2.5	0.06
Secondary outcome			
Length of stay, median (days)	11.1	12.9	1.00
30-day readmission, n (%)	0	1 (20%)	0.29
Inpatient mortality, n (%)	0	1 (20%)	0.29





Impact of Results on Patient Management

Oncology patient

- persistent spiking fevers
- received broad spectrum antibiotics; port in place
- all blood cultures negative; spiking fevers continued
- plan was to remove the port and hope the offending organism had been treated
- T2 positive for C. albicans/tropicalis
- port removed, Rx DC'd, placed on targeted antifungal therapy
- fever terminated; patient discharged



Summary

- Met Criteria of Performance
- Available 24/7
- Endorsed by "key" providers and service
- Impact study (expanded) on patient care, management, and antimicrobial use planned (Partnership with Pharmacy, Microbiology Laboratory, and Providers)
- A definite "TEAM" Effort
- Currently, the "only game in town"



THANK YOU!!!!

QUESTIONS/COMMENTS?

